

# A novel method for high-throughput analysis of bovine milk oligosaccharides

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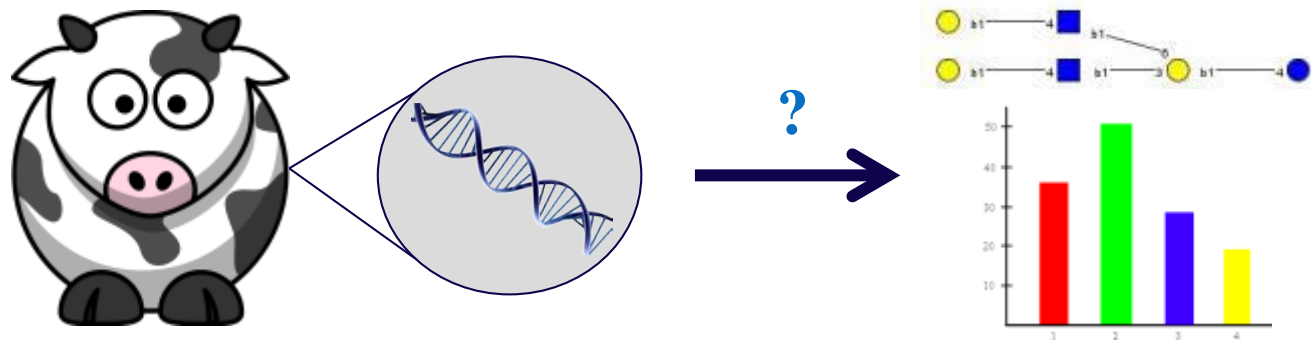
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# Project background

- DNA does not directly code for OS production, but it likely affects milk OS content indirectly



- Identifying genes that influence OS production would allow for improved OS utilization in dairy products

# Project background

- **The Danish-Swedish Milk Genomics Initiative aims to correlate relevant attributes of milk with the bovine genome to improve milk quality**
- **OS profiling and relative quantification will be conducted for 336 Holstein and 300 Jersey samples**
- **Relative OS quantities will be correlated with the genomes of the cows in a genome-wide association study**



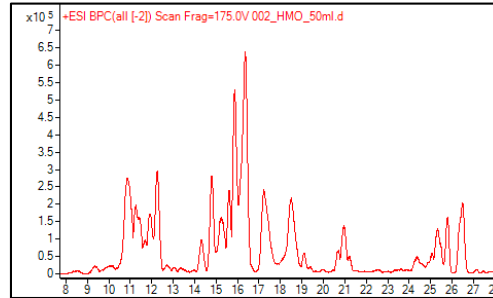
[milkgenomics.dk](http://milkgenomics.dk)

# Instrumental platform

- nano-HPLC separation with a porous graphitized carbon column
- Agilent 6520 Quadrupole time-of-flight (Q-TOF) mass spectrometer



# The throughput problem



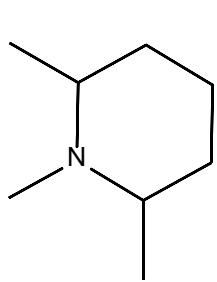
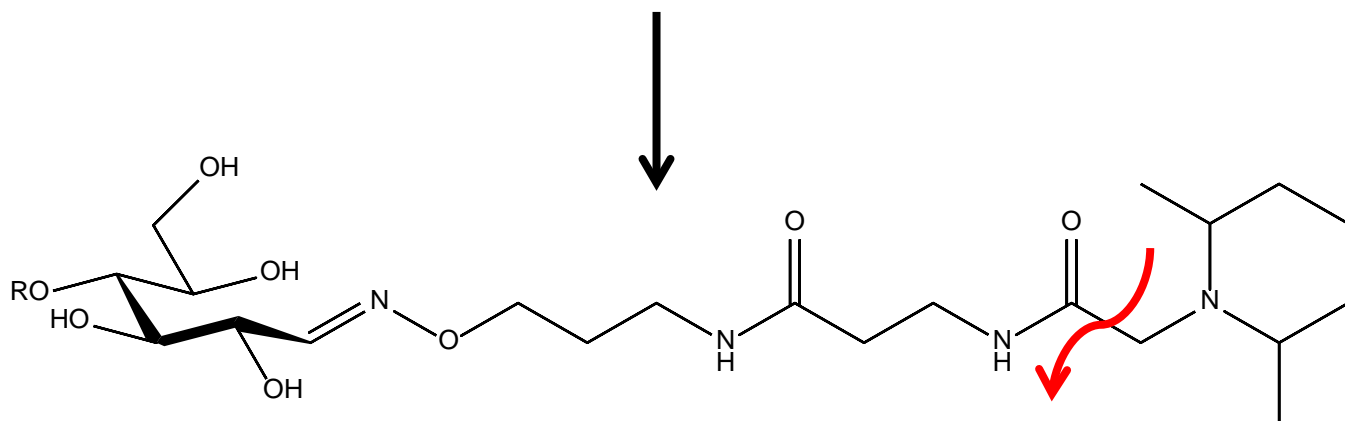
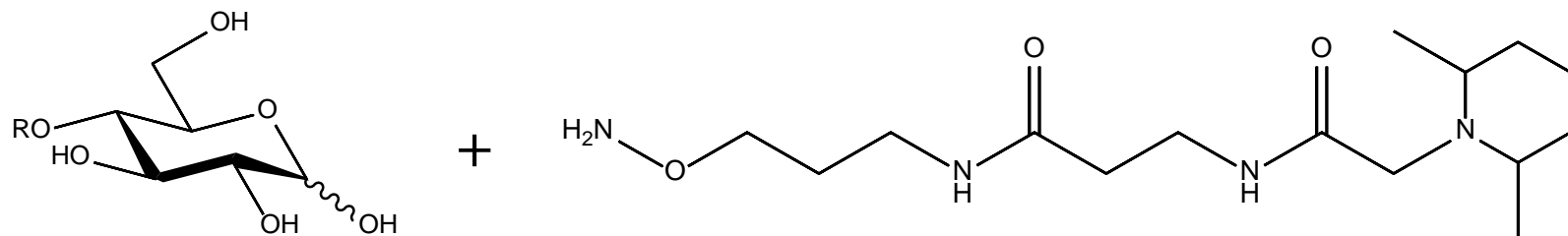
636 Samples X 70 min/sample

= 31 days

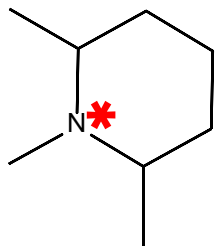


- + Blanks
- + Check standards
- + Tuning/calibration
- + Cleaning
- + Replicates

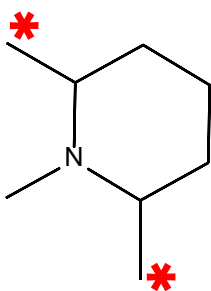
# Tandem mass tags



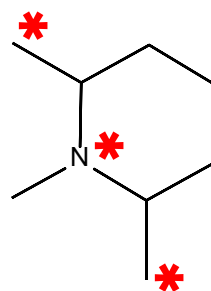
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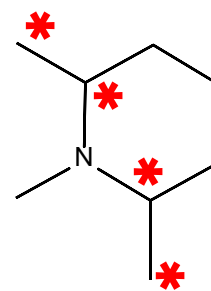
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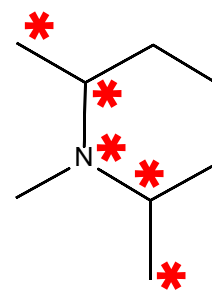
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129

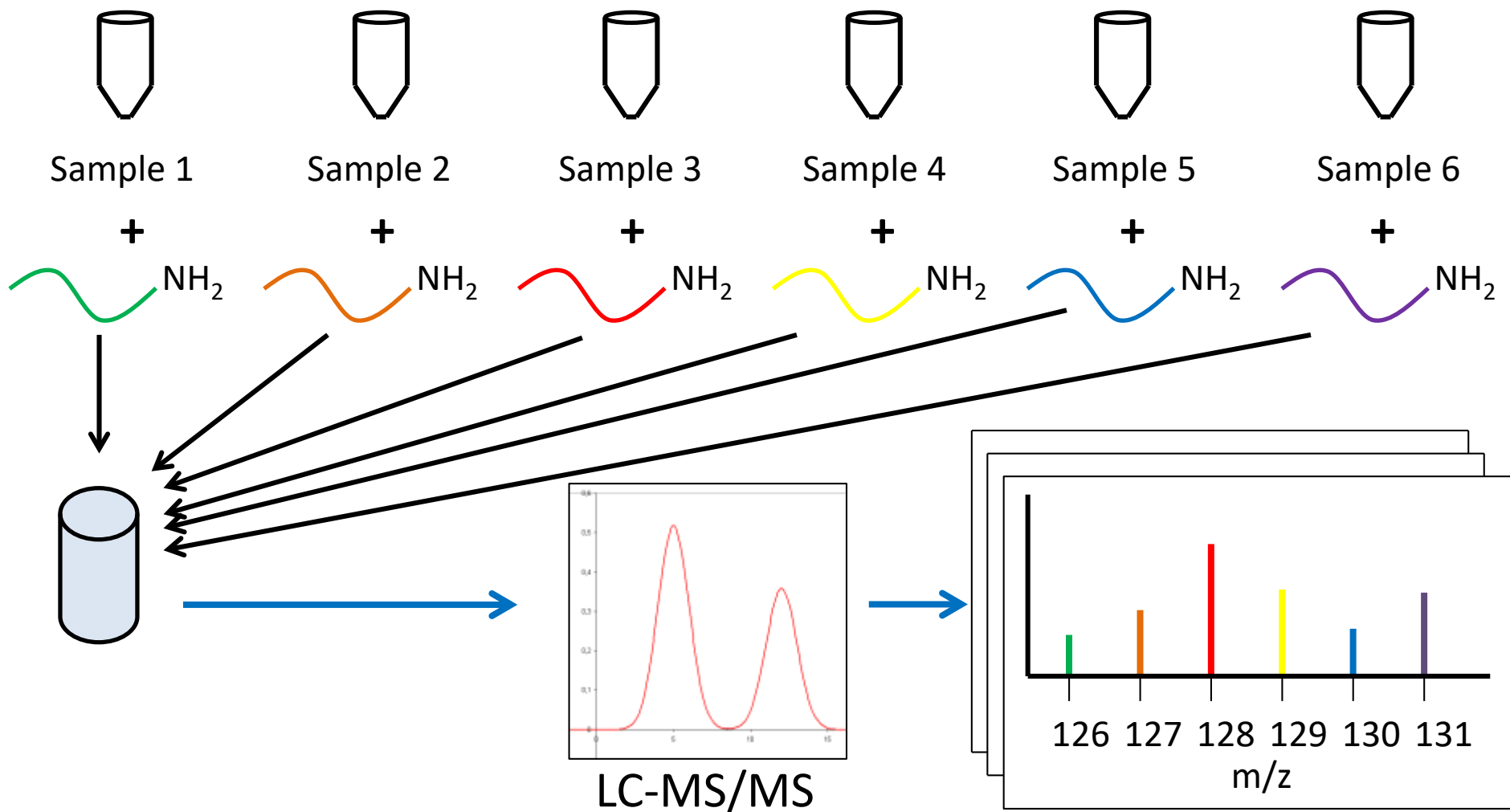


130



131

# Multiplexed analysis



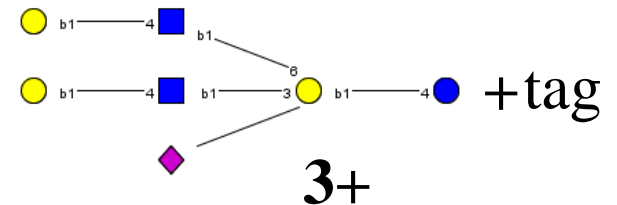
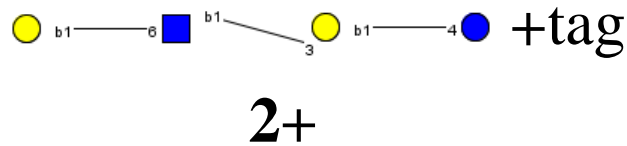
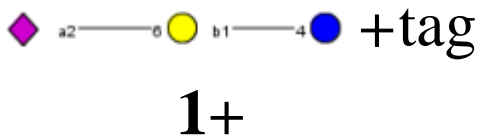
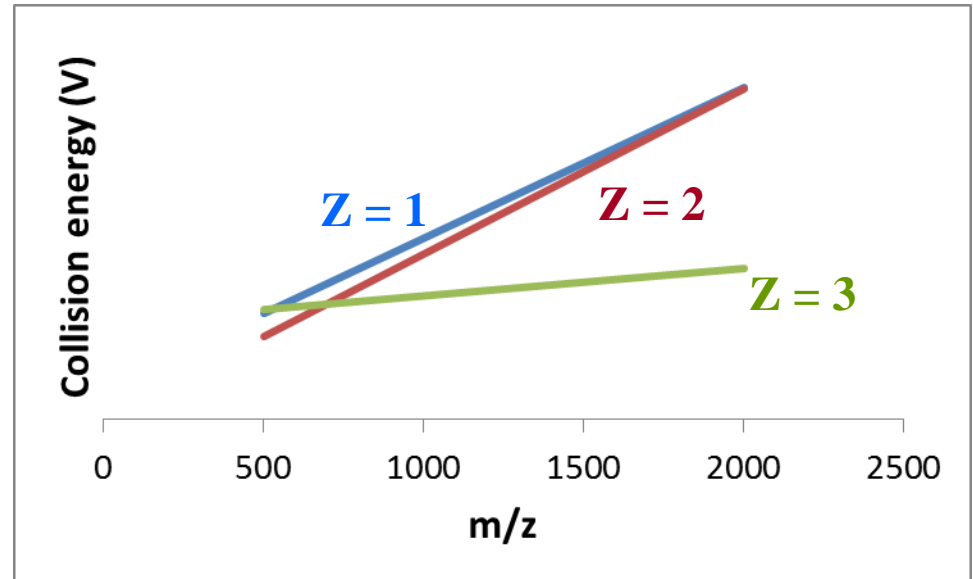
# Advantages

- **Reduction in instrumental runtime**
- **Less opportunity for drifts in sensitivity/performance**
- **Allows more samples to be run on a single column**
- **No manual peak integration**
- **Accurate relative quantification with optimized MS/MS method**

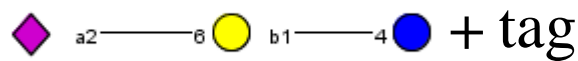
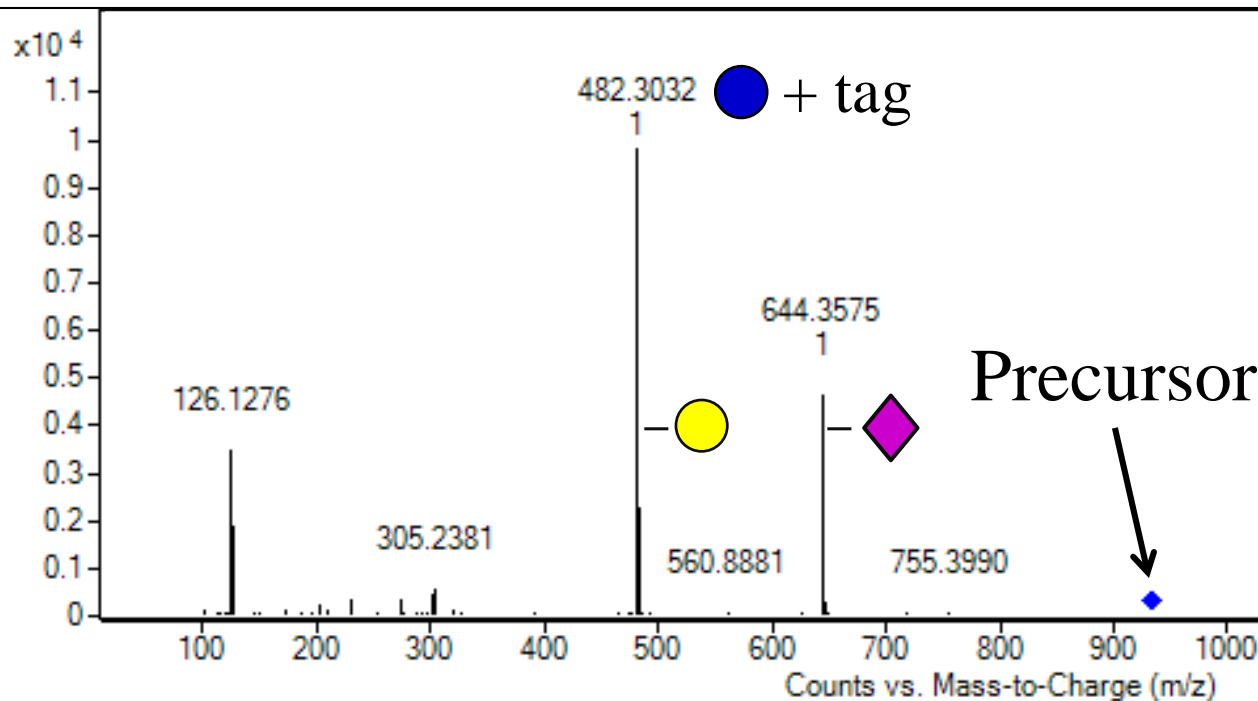


# Collision energies

- Data quality is highly dependent on collision energies
- Charge state influences fragmentation

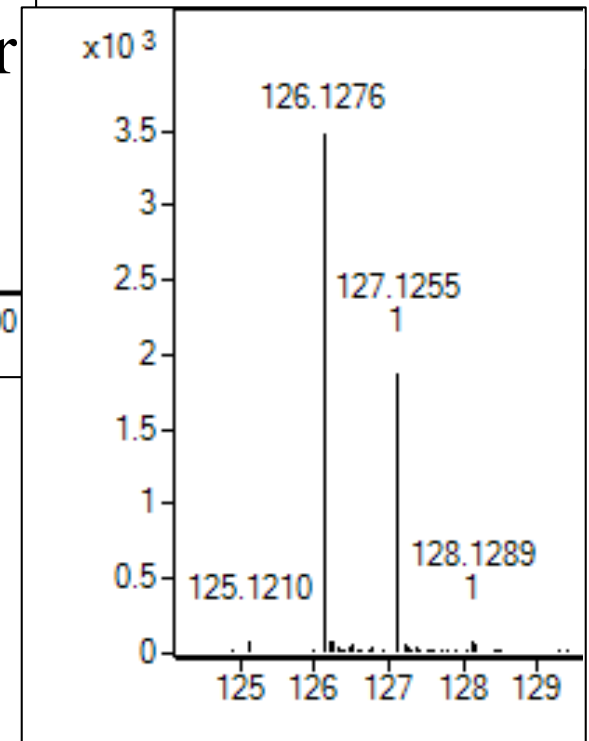


# Rapid & automated glycan IDs

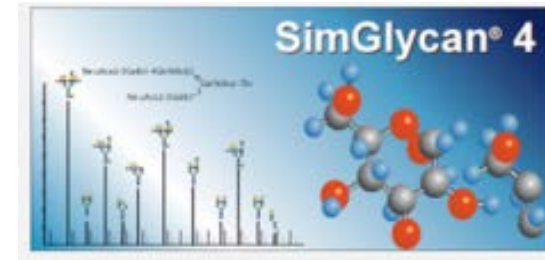
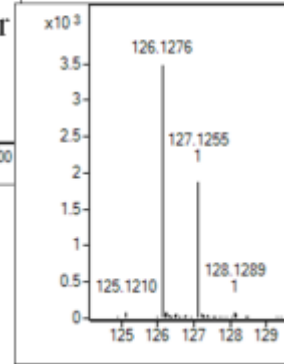
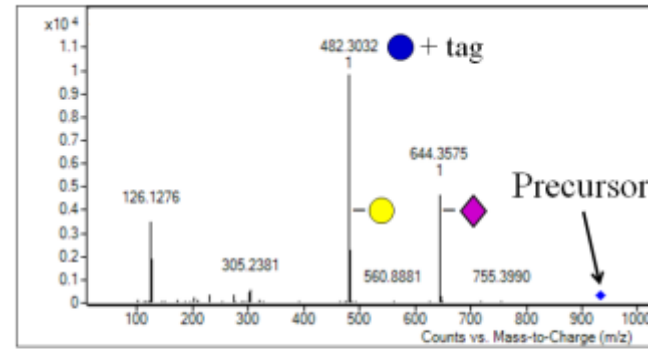
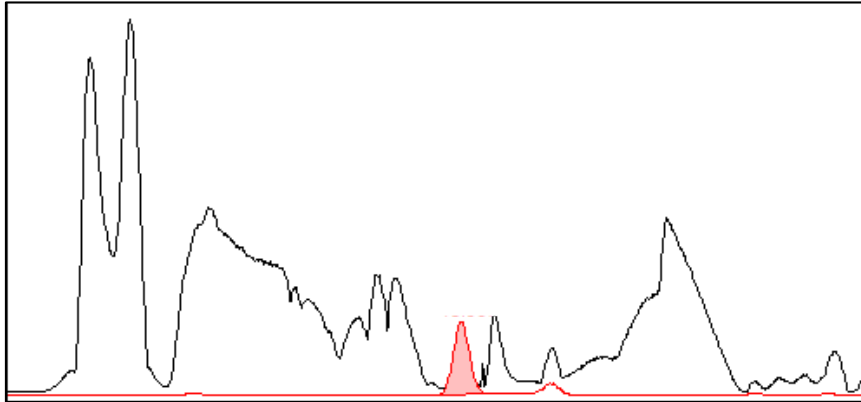


6'-sialyllactose

2 : 1



# Data analysis

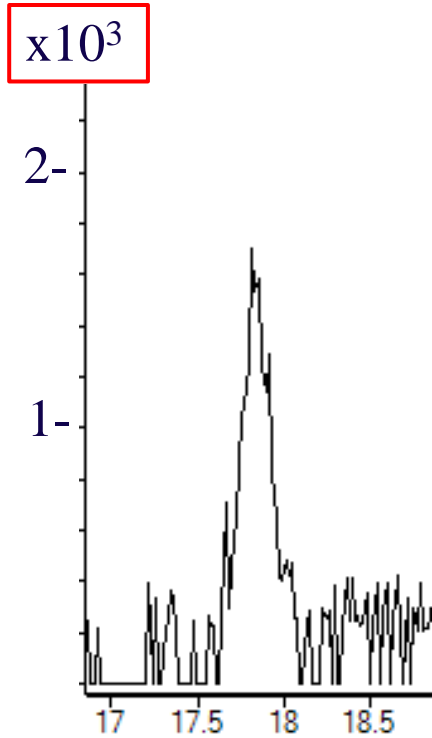


	A	B	C	D	E	F	G	H
	m/z	Composition	Mass	Ratio [TMT Ion/Control]	126.13	127.13	Glycan structure	
3	888.136093671	(Gal)1 (Glc)1 (NeuAc)1	633.211628077	1.0	0.671202247	1.0		
4	572.966922222	(Gal)1 (Glc)1 (NeuAc)1	633.211628077	1.0	0.479954764	1.0		
5	613.7788	(Hex)2 (NeuAc)2	924.307044605	1.0	1.225842485	1.0		
6	513.096678	(Hex)2 (HexNAc)1	545.195584082	1.0	0.377590916	1.0		
7	424.2222	(Gal)1 (Glc)1 (HexNAc)1	545.195584082	1.0	1.196550071	1.0		
8	549.2574	(Gal)2 (Glc)1 (NeuAc)1	795.264451509	1.0	0.642275801	1.0		
9	505.249	(Hex)3 (HexNAc)1	707.248407514	1.0	0.540409633	1.0		
10	606.7887	(Hex)3 (HexNAc)2	910.327780047	1.0	2.800464185	1.0		
11	708.3271	(Gal)3 (GlcNAc)1 (HexNAc)2	1113.40715258	1.0	0.095189109	1.0		
12	724.3227	(Fuc)1 (Hex)3 (HexNAc)6	1866.703178988	1.0	0.650984536	1.0		
13	586.276	(Gal)3 (Glc)1 (GlcNAc)1	869.301230945	1.0	2.015516586	1.0		



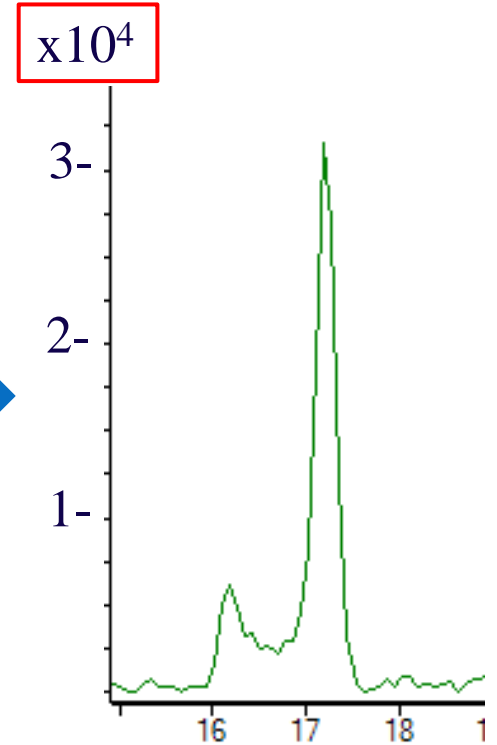
# Improved sensitivity

5 Hex  
4 HexNAc  
1 Fuc



Underivatized

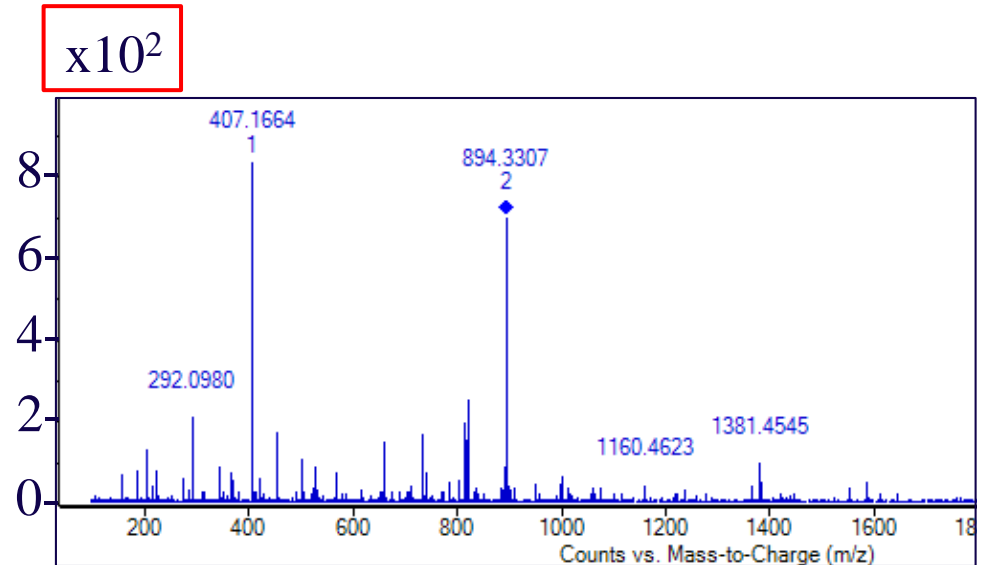
10X  
Increase



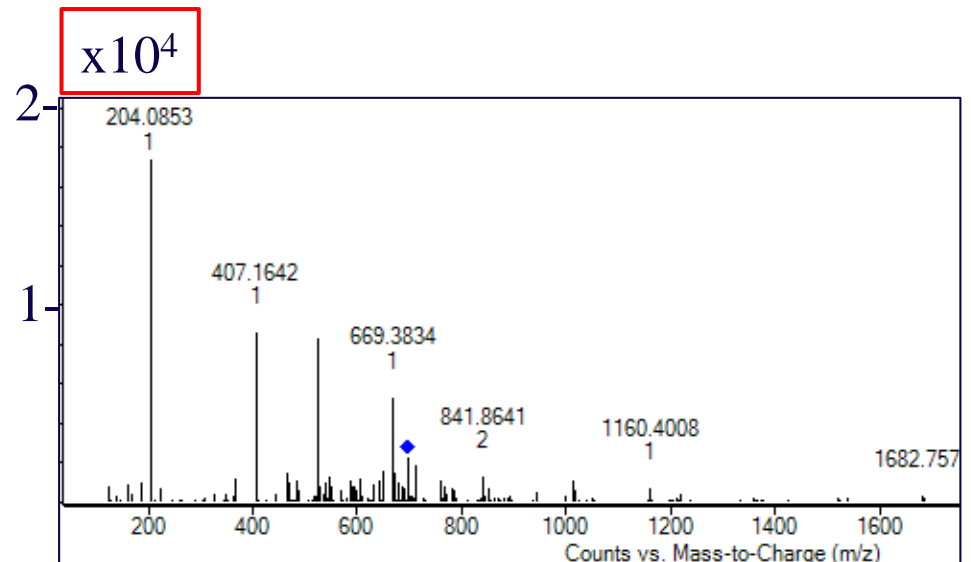
Aminoxy TMT-  
labeled sample

# Improved sensitivity

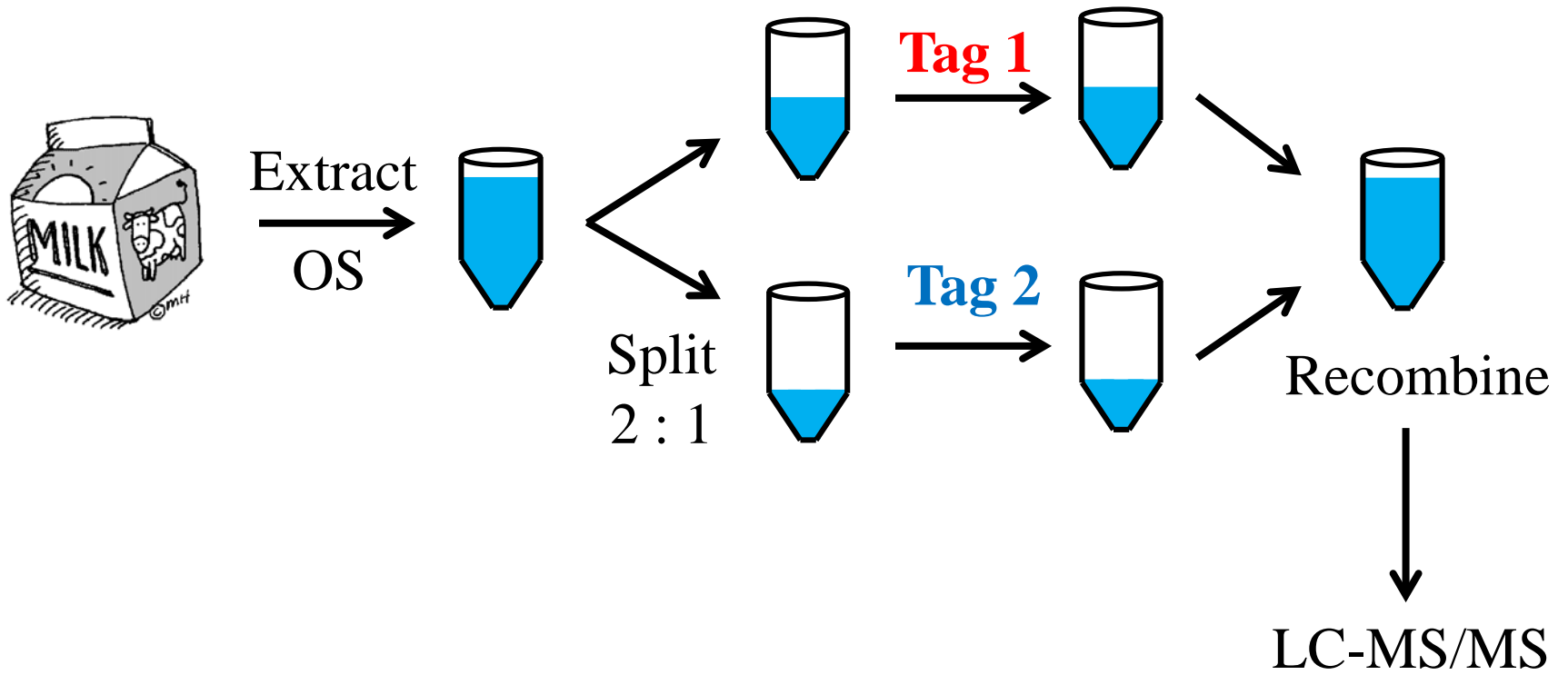
Underivatized



Aminoxy TMT-labeled sample



# Test with milk



# Acknowledgements



Daniela Barile



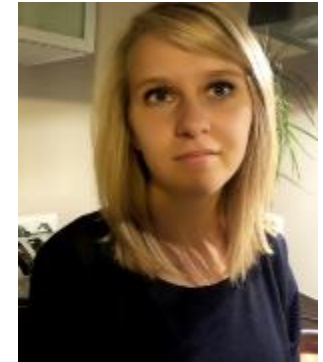
Lotte Bach  
Larsen



Nina Poulsen



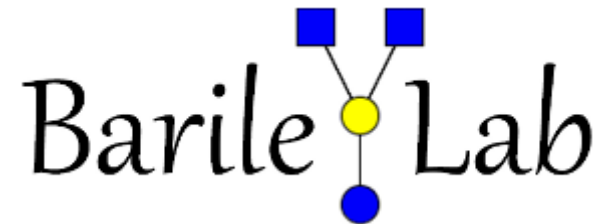
Emeline Colet



Chloe Duchene



National Institutes  
of Health



Daniela Barile

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